Tetrahedron Letters 51 (2010) 6153–6155

Contents lists available at ScienceDirect

Tetrahedron Letters

journal homepage: www.elsevier.com/locate/tetlet

The first total synthesis of the (±)-17-methyl-*trans*-4,5-methyleneoctadecanoic acid and related analogs with antileishmanial activity

Néstor M. Carballeira [a,*], Nashbly Montano [a], Rosa M. Reguera [b], Rafael Balaña-Fouce [b]

[a] Department of Chemistry, University of Puerto Rico, PO Box 23346, San Juan, PR 00931-3346, United States
[b] Department of Biomedical Sciences, University of Leon, Campus de Vegazana s/n, 24071 Leon, Spain

ARTICLE INFO

Article history:
Received 19 August 2010
Revised 16 September 2010
Accepted 17 September 2010

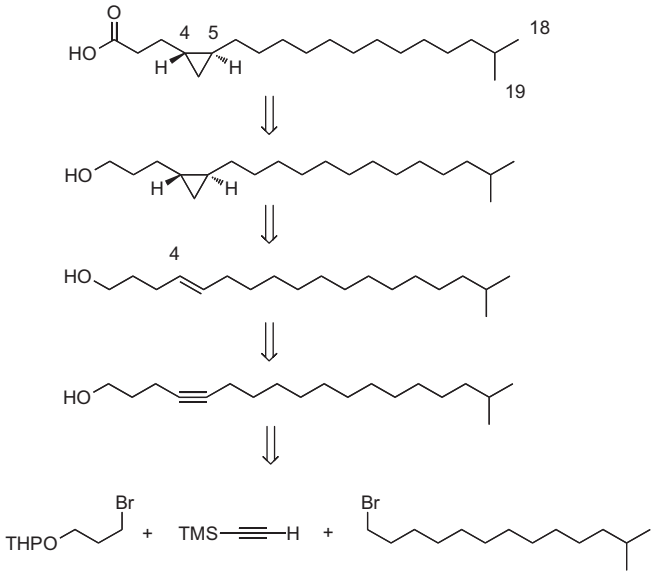
Keywords:
Antileishmanial activity
Cyclopropane fatty acids
Sponges
Synthesis

ABSTRACT

The first total synthesis of the marine cyclopropane fatty acid (±)-17-methyl-*trans*-4,5-methyleneoctadecanoic acid was accomplished in eight steps and in 9.1% overall yield starting from 1-bromo-12-methyltridecane. The *cis* analog (±)-17-methyl-*cis*-4,5-methyleneoctadecanoic acid was also synthesized but in seven steps and in 16.4% overall yield. With the two isomeric cyclopropane fatty acids at hand it was possible to unequivocally corroborate the *trans* relative configuration of the naturally occurring fatty acid by gas chromatographic co-elution of the corresponding methyl esters. The *cis* isomer was cytotoxic to *Leishmania donovani* promastigotes with an IC_{50} of 300.2 ± 4.2 μM.

Cyclopropane fatty acids (CFAs) are widespread in nature and they have been identified in many organisms ranging from bacteria to seed oils.[1] The earliest known example is lactobacillic acid (*cis*-11,12-methyleneoctadecanoic acid) but several structural variants have been isolated since.[1] One interesting compound is the 17-methyl-*cis*-9,10-methyleneoctadecanoic acid, from the protozoan *Herpetomonas megaseliae*, which incorporates both methyl and cyclopropyl branching in the chain.[2] While most of the known CFAs incorporate a *cis* cyclopropyl group in the acyl chain, just a few *trans* CFAs are known, such as the recently discovered 17-methyl-*trans*-4,5-methyleneoctadecanoic acid (**1a**) and the 18-methyl-*trans*-4,5-methylenenonadecanoic acid, which were identified in the phospholipids of the Caribbean sponge *Pseudospongosorites suberitoides*.[3] These marine fatty acids are quite interesting since they incorporate an unusual *trans* 4,5-cyclopropane in addition to *iso* methyl branching. However, the characterization of **1a** in the sponge extract was done by gas chromatography–mass spectrometry on suitable volatile derivatives followed by ^{1}H NMR of the total mixture of fatty acids. Therefore, a more rigorous confirmation of the structure of **1a** is warranted. For this purpose, a total synthesis of **1a** would not only serve to confirm the unusual *trans* cyclopropyl arrangement of the natural fatty acid, but also to report the total characterization of **1a**, as well as to provide the necessary expertise to synthesize analogs for biological screening. Therefore, herein we report the first total synthesis of both the naturally occurring (±)-17-methyl-*trans*-4,5-methyleneoctadecanoic acid (**1a**) and the corresponding *cis* analog **1b** together with the first studies of the antileishmanial activity of these CFAs.

A retrosynthetic analysis aimed at the synthesis of **1a** is outlined is Scheme 1. The *trans* cyclopropane fatty acid was envisioned as arising



Scheme 1. Retrosynthetic analysis toward the (±)-17-methyl-*trans*-4,5-methyleneoctadecanoic acid.

* Corresponding author. Tel.: +1 787 764 0000x4791; fax: +1 787 756 8242.
E-mail address: nmcarballeira@uprrp.edu (N.M. Carballeira).

0040-4039/$ - see front matter © 2010 Elsevier Ltd. All rights reserved.
doi:10.1016/j.tetlet.2010.09.083

from a *trans* olefin with the right chain length via a Simmons–Smith reaction.[4] The *trans* olefin, on the other hand, can be made from the corresponding alkyne using the standard sodium (Na) in ammonia (NH_3) reduction. A more elaborate construction is expected to be the introduction of the *iso* functionality in **1a** by means of the 1-bromo-12-methyltridecane, but the latter compound has been synthesized before in two steps starting from 2-bromopropane.[5]

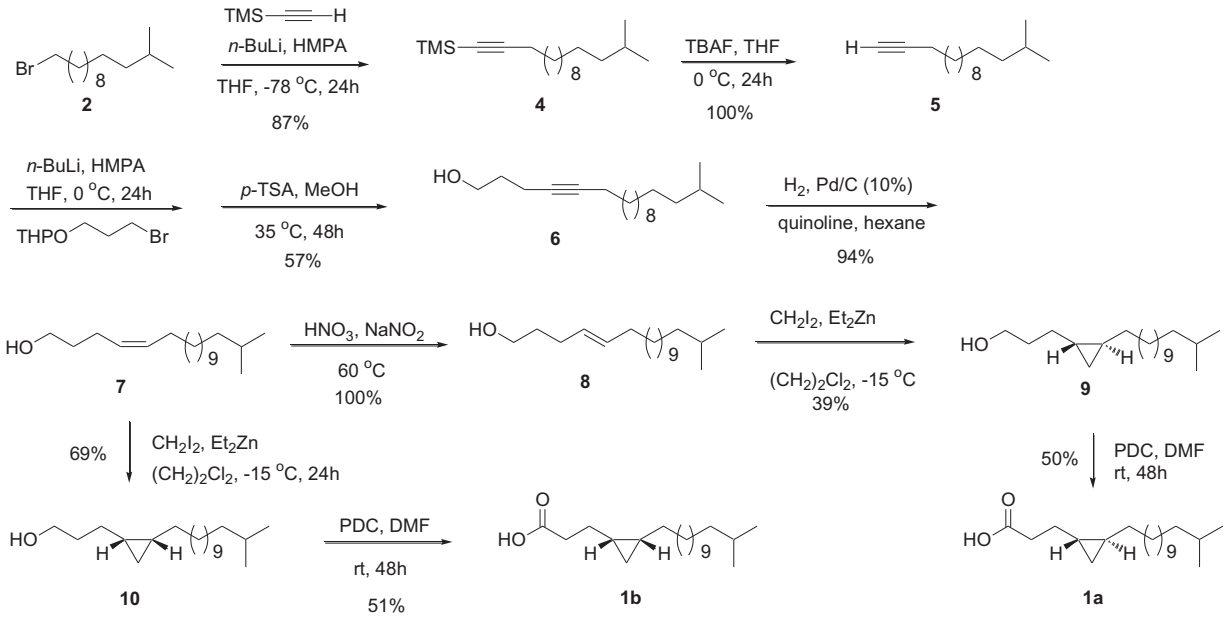
Our synthesis for the (±)-17-methyl-*trans*-4,5-methyleneoctadecanoic acid (**1a**) from the known 1-bromo-12-methyltridecane (**2**) is shown in Scheme 2. The first part of the synthesis called for the preparation of the key intermediate 17-methyloctadec-4-yn-1-ol (**6**), which can serve as a precursor for both the *trans* and *cis* cyclopropane fatty acids **1a** and **1b** (Scheme 2). For the introduction of the unsaturation at C-4, the (trimethylsilyl)acetylene was used and it was coupled to **2** using *n*-BuLi in THF-HMPA at −78 °C resulting in trimethyl(14-methylpentadec-1-ynyl)silane **4** in 87% yield. Desilylation of **4** with TBAF in THF at 0 °C yielded, in an almost quantitative yield, 14-methylpentadec-1-yne (**5**). The next step was the introduction of the precursor of the carboxy group at C-1 by coupling **5** with 2-(3-bromopropyloxy)-tetrahydro-2*H*-pyran by using *n*-BuLi in THF-HMPA at 0 °C (higher temperature for solubility reasons). In a subsequent step the tetrahydropyranyl group was removed using the standard procedure by adding catalytic amounts of *p*-TSA in methanol at 35 °C for 48 h, which yielded the desired 17-methyloctadec-4-yn-1-ol (**6**) in 57% yield for the two latter steps.

The final steps of the synthetic plan required using the alkyne in **6** to introduce both the *trans* and *cis* double bonds needed for the synthesis of the cyclopropanes **1a** and **1b**. Initially, the transformation of **6** into the (*E*)-17-methyloctadec-4-en-1-ol (**8**) was attempted with the classical dissolving metal reduction conditions of Na in liquid NH_3. However, all attempts to effectively carry out this transformation resulted in the partial conversion of **6** into **8**, probably due to the long alkyl chains. Failure to achieve a 100% reduction of **6** resulted in the need to effect a very difficult chromatographic separation of **6** and **8**, which was not practical. It was then decided to take a different route. Compound **6** was hydrogenated in hexane using H_2 under Lindlar catalysis, which afforded the (*Z*)-17-methyloctadec-4-en-1-ol (**7**) in 94% yield. The desired (*E*)-17-methyloctadec-4-en-1-ol (**8**) was effectively obtained by stereomutation of **7** with sodium nitrite–nitric acid in water at 60 °C.[6] This stereomutation worked quite well for this substrate and resulted in a quantitative yield of **8** from **7**. Alkenol **7** will also be used to prepare the corresponding *cis* cyclopropane fatty acid **1b**.

With the needed alkenols **7** and **8** at hand the cyclopropane ring was incorporated into the acyl chain by using the Simmons–Smith protocol, that is, diethyl zinc and diiodomethane in 1,2-dichloroethane under an argon atmosphere at −15 °C.[4] Under these conditions the 17-methyl-*trans*-4,5-methyleneoctadecan-1-ol (**9**) was obtained in 39% yield from **8**. The low yield in this reaction was due to the side-reaction of methylation of the alcohol resulting in the undesired methoxylated product. Attempts to protect the alcohol functionality in **8** with silyl protecting groups resulted in no reaction or very low yields of cyclopropanation. However, enough material of **9** was obtained by direct cyclopropanation of **8** to pursue the synthetic plan further. Final oxidation of **9** with pyridinium dichromate (PDC) in dimethylformamide (DMF) under an argon atmosphere resulted in 50% yield of the desired *trans* acid **1a**.[7] Identical conditions were also used to obtain **1b** from **7**. This means that cyclopropanation of **7** under the same Simmons–Smith conditions described above resulted in 69% yield of the 17-methyl-*cis*-4,5-methyleneoctadecan-1-ol (**10**) and further oxidation to the acid with PDC in DMF yielded the expected (±)-17-methyl-*cis*-4,5-methyleneoctadecanoic acid (**1b**) in 51% yield.[8]

With both acids **1a** and **1b** at hand we were in a good position to unequivocally corroborate the relative *trans* cyclopropane stereochemistry as well as the structure of the natural fatty acid **1a** that was assigned on the basis of ^{1}H NMR spectroscopy on the whole fatty acid mixture from the sponge *P. suberitoides*.[3] This was done by gas chromatographic co-injection of the corresponding methyl esters of **1a** and **1b**, prepared from the acids by esterification with MeOH and catalytic amounts of HCl, with the fatty acid methyl ester mixture from the phospholipids of the sponge *P. suberitoides*.[3] In this experiment the methyl ester of synthetic **1a** co-eluted (in a HP-5MS capillary column) with the natural cyclopropane methyl ester (ECL = 19.15), thus unequivocally confirming the structure of the natural fatty acid as well as its *trans* 4,5-cyclopropane stereochemistry.

We had previously shown that the *iso* methyl-branched monounsaturated fatty acid (*Z*)-17-methyl-13-octadecenoic acid displays antileishmanial activity toward *Leishmania donovani* promastigotes



Scheme 2. Synthesis of the (±)-17-methyl-*trans*-4,5-methyleneoctadecanoic acid (**1a**) and the (±)-17-methyl-*cis*-4,5-methyleneoctadecanoic acid (**1b**).

with an EC_{50} = 19.8 ± 7.0 μg/ml and, as a probable intramolecular target, inhibits the leishmania DNA topoisomerase IB enzyme at concentrations of 50 μM.[9] Given these previous results we decided to test the *cis* cyclopropane fatty acid **1b** against *L. donovani* promastigotes and establish how cyclopropane substitution compares to monounsaturation in determining the antileishmanial activity of these *iso*-C_{18} fatty acids.[10] It was found that acid **1b** was cytotoxic to *L. donovani* promastigotes at an IC_{50} = 300.2 ± 4.2 μM and it did not inhibit the leishmania DNA topoisomerase IB enzyme. Therefore, monounsaturation is more effective than cyclopropanation with respect to increasing the cytotoxicity of these *iso*-C_{18} fatty acids toward *L. donovani*. It is important to mention that the chain length also plays a role in the antileishmanial activity of these CFAs. The longer chain analog (±)-18-methyl-*cis*-4,5-methylenenonadecanoic acid, also synthesized by us following a similar route as that described in Scheme 2, displayed no activity against the *L. donovani* promastigotes (IC_{50} >1000 μM). Therefore, other shorter chain analogs could be synthesized in order to find the optimum chain length for antileishmanial activity. The synthetic route reported herein will facilitate the preparation of these analogs.

Acknowledgments

The project described was supported by Award Number SC1GM084708 from the National Institutes of General Medical Sciences of the NIH. We thank Dr. Fred Strobel (Emory University) for the high resolution mass spectral data. This research was also partially supported by a grant (Gr238) from Junta de Castilla y León, PS09/00448, and the Tropical Diseases Network (RICET) from Ministerio de Salud y Consumo (SPAIN).

References and notes

1. (a) Grogan, D. W.; Cronan, J. E., Jr. *Microbiol. Mol. Biol. Rev.* **1997**, *61*, 429; (b) Fish, W. R.; Holz, G. G., Jr.; Beach, D. H.; Owen, E.; Anekwe, G. E. *Mol. Biochem. Parasitol.* **1981**, *3*, 103; (c) Gontier, E.; Boussouel, N.; Terrasse, C.; Jannoyer, M.; Menard, M.; Thomasset, B.; Bourgaud, F. *Biochem. Soc. Trans.* **2000**, *28*, 578; (d) Cox, A. D.; Wilkinson, S. G. *Biochim. Biophys. Acta* **1989**, *1001*, 60.
2. Holz, G. G., Jr.; Beach, D. H.; Singh, B. N.; Fish, W. R. *Lipids* **1983**, *18*, 607.
3. Carballeira, N. M.; Montano, N.; Vicente, J.; Rodriguez, A. D. *Lipids* **2007**, *42*, 519.
4. (a) Simmons, H. E.; Smith, R. D. *J. Am. Chem. Soc.* **1958**, *80*, 5323; (b) Simmons, H. E.; Smith, R. D. *J. Am. Chem. Soc.* **1959**, *81*, 4256.
5. Mun, J. Y.; Onorato, A.; Nichols, F. C.; Morton, M. D.; Saleh, A. I.; Welzel, M.; Smith, M. B. *Org. Biomol. Chem.* **2007**, *5*, 3826.
6. (a) Bumpus, F. M.; Taylor, W. R.; Strong, F. M. *J. Am. Chem. Soc.* **1950**, *72*, 2116; (b) Duffy, P. E.; Quinn, S. M.; Roche, H. M.; Evans, P. *Tetrahedron* **2006**, *62*, 4838.
7. Spectral data for the (±)-17-methyl-*trans*-4,5-methyleneoctadecanoic acid (**1a**): transparent oil, IR (neat) ν_{max} 3500–2500, 2923, 2853, 1711 (C=O), 1464, 1383, 1365, 1274, 1120, 1073, 1039, 737 cm^{-1}; ^1H NMR ($CDCl_3$, 300 MHz) δ 2.42 (2H, t, J = 7.4 Hz, H-2), 1.65–1.51 (3H, m, H-3, H-17), 1.27 (21H, m, $-CH_2-$), 1.16 (2H, m, $-CH_2-$, H-16), 0.86 (6H, d, J = 6.6 Hz, H-18, H-19), 0.44 (2H, m, H-4, H-5), 0.21 (2H, t, J = 6.1 Hz, CH_2 in cp ring); ^{13}C NMR ($CDCl_3$, 75 MHz) δ 177.63 (s, C-1), 39.04 (t, C-16), 34.09 (t, C-6), 30.33 (t, C-2), 29.94 (t), 29.68 (t), 29.55 (t), 29.51 (t), 29.90 (t), 27.95 (d, C-17), 27.41 (t), 25.43 (t), 22.65 (q, C-18, C-19), 18.90 (d, C-4), 18.06 (d, C-5), 11.78 (t, CH_2 in cp ring). HRMS (APCI): calcd for $C_{20}H_{37}O_2$ [M^+-1] 309.2799, found 309.2798.
8. Spectral data for the (±)-17-methyl-*cis*-4,5-methyleneoctadecanoic acid (**1b**): transparent oil, IR (neat) ν_{max} 3500–2500, 2922, 2852, 1709 (C=O), 1459, 1382, 1365, 1274, 1078, 1039, 721 cm^{-1}; ^1H NMR ($CDCl_3$, 300 MHz) δ 2.45 (2H, t, J = 7.6 Hz, H-2), 1.71 (1H, m, H-18), 1.51 (2H, m, H-3), 1.26 (21H, m, $-CH_2-$), 1.15 (2H, m, $-CH_2-$, H-16), 0.86 (6H, d, J = 6.6 Hz, H-18, H-19), 0.71 (2H, m, H-4, H-5), 0.60 (1H, m, one CH_2 in cp ring), −0.26 (1H, m, one CH_2 in cp ring); ^{13}C NMR ($CDCl_3$, 75 MHz) δ 179.19 (s, C-1), 39.05 (t, C-16), 34.50 (t, C-6), 30.33 (t, C-2), 30.15 (t, C-3), 29.94 (t), 29.78 (t), 29.70 (t), 29.68 (t), 28.57 (t), 27.96 (d, C-17), 27.42 (t, C-7), 24.10 (t), 22.66 (q, C-18, C-19), 15.97 (d, C-4), 15.10 (d, C-5), 10.77 (t, CH_2 in cp ring). HRMS (APCI): calcd for $C_{20}H_{37}O_2$ [M^+-1] 309.2799, found 309.2798.
9. Carballeira, N. M.; Montano, N.; Balaña-Fouce, R.; Fernández Prada, C. *Chem. Phys. Lipids* **2009**, *161*, 38.
10. For experimental details on the antileishmanial testing on *L. donovani* (MHOM/ET67/L82 strain) promastigotes see Ref. 9 above.